### Note

# Synthesis of reducing disaccharides bearing a lipophilic chain for the conjugation to proteins\*

DANIEL CABARET, RAFI KAZANDJAN, AND MICHEL WAKSELMANT

Centre d'Étude et de Recherche de Chimie Organique Appliquée, CNRS, 2 rue Henry Dunant, F-94320 Thiais (France)

(Received September 16th, 1985; accepted for publication, November 18th, 1985)

Some native or chemically modified enzymes are active in non-aqueous solvents<sup>1,2</sup>. However, the use of enzymes in organic synthesis is generally limited by thermal and solvent-induced denaturations<sup>3-6</sup>.

In mixtures of water and class I solvents (polyols, sugars, etc.), preferential hydration of the proteins occurs and leads to stabilization<sup>7</sup>. Some nonionic and nondenaturing detergents, such as alkyl  $\beta$ -D-glucopyranosides or N-(D-gluco)-N-methylalkanamides are used for the solubilization of membranes<sup>8,9</sup>. Thus, our goal was to synthesize reducing disaccharides bearing an hydrophobic chain (A) and to conjugate them with lysine residues of proteins<sup>10</sup>, so as to obtain new neoglycoproteins<sup>11</sup>. In organic media having low proportions of water, these conjugates might have the structure B shown in Scheme 1. A few layers of water, hydrogenbonded to the polyol chains, could protect the proteins from the solvent in which the hydrophobic chain is extended.

Two different disaccharides of type A have been prepared by following the oxazoline or the modified Koenigs-Knorr procedure of glycosylation<sup>12,13</sup>. The first

(HO)<sub>n</sub> O 
$$CH_2$$
  $CH_2$   $CH_2$ 

Scheme 1

<sup>\*</sup>Presented at the Third European Symposium on Carbohydrates, Grenoble, September 16–20, 1985. \*To whom correspondence should be addressed.

$$ACOCH_{2}$$

$$ACOC$$

has a long-chain amido substituent, the second a long-chain ether group.

Acylation of 2-amino-2-deoxy-D-glucose with decanoyl chloride gave the known 2-decanamido-2-deoxy-D-glucose. Treatment of this amide with acetyl chloride saturated with dry hydrogen chloride yielded a crude acylated glycosyl chloride<sup>14</sup> which cyclized in the presence of silver nitrate and 2,4,6-trimethyl-pyridine<sup>15</sup> to give the long-chain oxazoline 1 in 25% overall-yield. Condensation of 1 with 1,2:3,4-di-O-isopropylidene-D-galactopyranose (2) in the presence of trifluoromethanesulfonic acid<sup>16</sup> led to the protected disaccharide 3 in 54% yield. The cleavages of the acetal groups with 90% trifluoroacetic acid<sup>17</sup> and of acetate groups with methanol-triethylamine gave O-(2-decanamido-2-deoxy- $\beta$ -D-glucopyranosyl)-(1 $\rightarrow$ 6)-D-galactopyranose 5 in 80% yield.

Alkylation of 2 with octyl bromide in N, N-dimethylformamide containing sodium hydride<sup>18</sup> gave the ether 6 in a 68% yield, and cleavage of the ketal groups the 6-O-octyl-D-galactopyranose 7. An homologous dodecylated surfactant has been prepared under more drastic conditions by an analogous sequence of reactions<sup>19</sup>. Acetylation of 7 and treatment of the resulting tetraacetate with hydrogen bromide led to a glycosyl bromide 8 having a neigboring ester group. Condensation of 8 with 2 in the presence of silver trifluoromethanesulfonate and 1,1,3,3-tetramethylurea<sup>20</sup> gave a mixture from which the protected disaccharide 9 was isolated

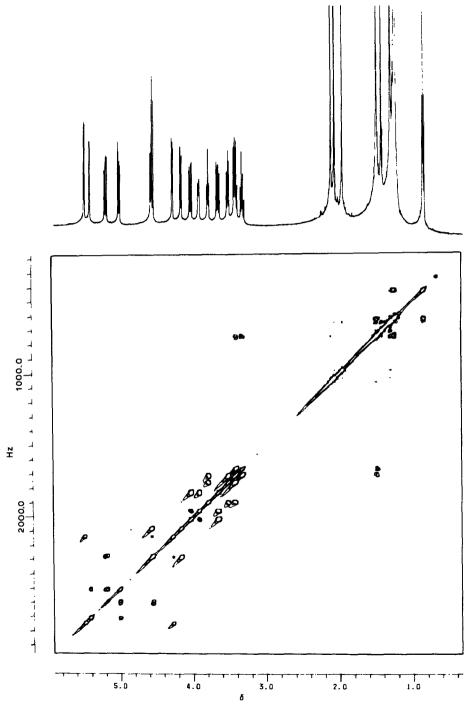


Fig. 1. Contour plot of the 2D-COSY n.m.r. spectrum of 9 in CDCl<sub>3</sub>. The various cross peaks on either side of the diagonal arose owing to the presence of J couplings between various spin multiplets shown in the correpsonding 1D-n.m.r. at the top.

in 40% yield by chromatography. Its structure was confirmed by 2D-COSY n.m.r. spectroscopy<sup>21</sup> (Fig. 1). Cleavages of the protecting groups led to O-(6-O-octyl- $\beta$ -D-galactopyranosyl)-(1 $\rightarrow$ 6)-D-galactopyranose (10).

#### **EXPERIMENTAL**

General methods. — Melting points were determined with a Mettler FP61 apparatus. <sup>1</sup>H-N.m.r. spectra were recorded at 90 or 500 MHz, <sup>13</sup>C-n.m.r. spectra at 20 MHz; unless otherwise stated, the solvent was CDCl<sub>3</sub>. T.l.c. were developed on Silica gel 60F (Merck) and spots detected with 10% H<sub>2</sub>SO<sub>4</sub> in water or with a solution of (NH<sub>4</sub>)<sub>2</sub>MoO<sub>4</sub> (5 g), H<sub>2</sub>SO<sub>4</sub> (5 mL), H<sub>3</sub>PO<sub>4</sub> (5 mL) in water (100 mL), after heating. Column chromatography were made on Silica gel Merck 60 (70–230 mesh). The compounds described subsequently were homogeneous on t.l.c.

2-Nonyl-(3,4,6-tri-O-acetyl-1,2-dideoxy-α-D-glucopyrano)-[2,1-d]-2-oxazoline (1). — A solution of 2-decanamido-2-deoxy-D-glucose<sup>14</sup> (3.33 g, 10 mmol) in acetyl chloride (60 mL) was saturated with dry HCl at -15°. Then, the flask was stoppered and the mixture stirred for 24 h at room temperature. The solution was evaporated, the residue dissolved in dichloromethane (25 mL), quickly washed with cold water, and then with a cold saturated aqueous NaHCO<sub>3</sub> solution, dried (MgSO<sub>4</sub>), and the solvent evaporated. The crude chloride was dissolved in acetone (30 mL) and slowly added with stirring to a suspension of AgNO<sub>3</sub> (2 g) in acetone (30 mL) and 2,4,6-trimethylpyridine (5 mL) which was stirred for 2 h at room temperature. Chloroform (50 mL) was added, the suspension filtered, and the filtrate washed with an aqueous KHCO<sub>3</sub> solution and dried (K<sub>2</sub>CO<sub>3</sub>). Solvent removal in vacuo left a syrupy residue which was purified by column chromatography in 4:1 pentaneethyl acetate to give 1 (1.10 g, 25%);  $\nu_{\text{max}}^{\text{film}}$  1725 (OAc) and 1645 cm<sup>-1</sup> (C=N); <sup>1</sup>Hn.m.r. data:  $\delta$  6.0 (d,  $J_{1,2}$  7 Hz, H-1), 5.3 (t, H-3), 5.,0 (m, H-5), 4.1 (m, H-4 and -6), 3.6 (m, H-2), 2.4 (t, H $\alpha$ ), 2.2–2.1 (3 s, COCH<sub>3</sub>), 1.5 [m, (CH<sub>2</sub>)<sub>7</sub>], and 0.9 (t, CH<sub>3</sub>). Compound 1 was stable when kept at 4°, but was immediately used for glycosylation reactions.

1,2:3,4-Di-O-isopropylidene-6-O-(3,4,6-tri-O-acetyl-2-decanamido-2-deoxy-β-D-glucopyranosyl)-α-D-galactopyranose (3). — A solution of the oxazoline 1 (0.75 g, 1.7 mmol), 1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (2) (0.44 g, 1.7 mmol), and trifluoromethanesulfonic acid (10 μL) in dry dichloroethane (10 mL) was heated under an Ar atmosphere for 20 min at 80°, and then kept for 1 h at room temperature. The solution was washed with cold aqueous NaHCO<sub>3</sub> and dried (Na<sub>2</sub>SO<sub>4</sub>). The filtered solution was evaporated, and the residue applied to a silica gel column and eluted with 2:3 ethyl acetate-pentane to give 3 (0.66 g, 54%), m.p. 151.7°,  $[\alpha]_D^{20} - 38^\circ$ ,  $[\alpha]_{346}^{29} - 45^\circ$  (c 0.3, methanol); <sup>1</sup>H-n.m.r. δ; 5.7 (d,  $J_{1,2}$  5 Hz, H-1), 4.72 (d,  $J_{1',2'}$  9 Hz, H-1'), 2.2, 2.1 (s, COCH<sub>3</sub>), 1.52, 1.42, and 1.27 [s, C(CH<sub>3</sub>)<sub>2</sub>]; the remainder of the spectrum was not interpreted; <sup>13</sup>C-n.m.r.: δ 173.6 (CONH), 170.9, 169.6 (3 C, O-C=O), 109.6, 108.8 (2 C, O-C-O), 101.8 (C-1'), 96.4 (C-1), 73.2-62.3 (9 C, cyclic), 54.2 (C-2'), and 36.8-14.2 (16 C).

Anal. Calc. for  $C_{34}H_{55}NO_{14}$ : C, 58.1; H, 7.9; N, 2.0. Found: C, 57.8; H, 7.9; N, 2.0.

6-O-(2-Decanamido-2-deoxy-β-D-glucopyranosyl)-1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (4). — Compound 3 (700 mg, 1 mmol) was deacetylated by stirring for 20 h at room temperature with methanol (15 mL) and triethylamine (2 mL). The solution was evaporated and the residue purified by column chromatography (in 17:3 ethyl acetate-ethanol) to give 4 (520 mg, 90%), m.p.  $147^{\circ}$ ,  $[\alpha]_{20}^{20}$  -33°,  $[\alpha]_{546}^{20}$  -39° (c 0.3, methanol); <sup>1</sup>H-n.m.r.: δ 5.6 (d,  $J_{1,2}$  5 Hz, H-1), 1.53, 1.46, and 1.30 [s, C(CH<sub>3</sub>)<sub>2</sub>]; <sup>13</sup>C-n.m.r.: δ 176.9 (CONH), 110.2, 109.7 (2 C, O-C-O), 103.3 (C-1'), 97.6 (C-1), 78.0-62.9 (9 C, cyclic), 57.3 (C-2'), and 37.6-14.4 (remaining C).

Anal. Calc. for  $C_{23}H_{49}NO_{11} \cdot H_2O$ : C, 56.6; H, 8.6. Found: C, 56.6; H, 8.2.

6-O-(2-Decanamido-2-deoxy-β-D-glucopyranosyl)-D-galactopyranose (5). — Compound 4 (290 mg, 0.5 mmol) was dissolved in 90% aqueous trifluoroacetic acid (3 mL) and kept for 1 h at 0°. The solution was evaporated to give 225 mg (90%) of a chromatographically pure product. It crystallized from 90% aqueous methanol to give the disaccharide monohydrate, m.p. 179°,  $[\alpha]_D^{20} = -3.7^\circ$ ,  $[\alpha]_{546}^{20} +1.9^\circ$  (c 3, pyridine); <sup>13</sup>C-n.m.r. [(<sup>2</sup>H<sub>5</sub>)-pyridine]: δ 174.6 (CO), 103.0 (C-1'), 99.5 (Cβ-1), 94.5 (Cα-1), 78.5-62.7 (C, cyclic), 57.5 (C-2'), and 37.0-14.3 (remaining C).

Anal. Calc. for  $C_{22}H_{41}NO_{11} \cdot H_2O$ : C, 51.5; H, 8.4; N, 2.8. Found: C, 51.9; H, 8.2; N, 2.8.

1,2:3,4-Di-O-isopropylidene-6-O-octyl-α-D-galactopyranose (6). — A solution of 1,2:3,4-di-O-isopropylidene-α-D-galactopyranose (2) (2.6 g, 10 mmol) in N,N-dimethylformamide (15 mL) was slowly added to a suspension of NaH (480 mg, 20 mmol) in dry N,N-dimethylformamide (30 mL). After 1 h at room temperature, the suspension was cooled to 0° and bromooctane (2.3 g, 12 mmol) was slowly added. The mixture was stirred for 20 h at room temperature, methanol (5 mL) was added, and the solvent removed. The residue was applied to a silica gel column and eluted with 9:1 pentane—ethyl acetate to give 6 (2.56 g, 68%),  $[\alpha]_{\rm D^0}^{20}$  –48°,  $[\alpha]_{\rm 546}^{20}$  –56.6° (c 2, methanol); <sup>1</sup>H-n.m.r.: δ 5.55 (d,  $J_{1,2}$  5 Hz, H-1), 4.60 (m,  $J_{3,4}$  8 Hz, H-3), 4.30 (m,  $J_{2,3}$  2.3 Hz, H-2), 4.27 (m,  $J_{4,5}$  1.7 Hz, H-4), 4.0 (m,  $J_{5,6}$  6.3 Hz, H-5), 3.6 (d, H-6), 3.5 (t,  $J_{\alpha,\beta}$  6 Hz, H-α), 1.55, 1.46, 1.35 [3 s, C(CH<sub>3</sub>)<sub>2</sub>], 1.3 [m, (CH<sub>2</sub>)<sub>6</sub>], and 0.9 (t, CH<sub>3</sub>).

Anal. Calc. for C<sub>20</sub>H<sub>36</sub>O<sub>6</sub>: C, 64.5; H, 9.7. Found: C, 64.5; H, 9.6.

6-O-Octyl- $\alpha$ -D-galactopyranose (7). — Compound 6 (2.36 g) was dissolved in 90% aqueous trifluoroacetic acid (10 mL). The solution was kept for 1 h at 0°, and then evaporated to give a chromatographically pure product. Crystallization from ethanol gave 7, (1.5 g, 81%), m.p. 153°,  $[\alpha]_D^{20}$  +32.9°,  $[\alpha]_{546}^{20}$  +38.8° (c 1.1, methanol).

Anal. Calc. for C<sub>14</sub>H<sub>28</sub>O<sub>6</sub>: C, 57.5; H, 9.6. Found: C, 57.3; H, 9.6.

2,3,4-Tri-O-acetyl-6-O-octyl- $\alpha$ -D-galactopyranosyl bromide (8). — Compound 7 (2.9 g) was dissolved in acetic anhydride (30 mL) with BF<sub>3</sub> · Et<sub>2</sub>O (0.5 mL) as catalyst. After 3 h, at room temperature, the solvent was removed and the

residue dissolved in a saturated solution of HBr in acetic acid (15 mL). After 3 h at room temperature, HBr and acetic acid were eliminated under reduced pressure. The syrupy residue was purified by column chromatography in 1:4 diethyl etherpentane to give **8** (3.3 g, 70%);  $^{1}$ H-n.m.r.:  $\delta$  6.6 (d,  $J_{1,2}$  5 Hz, H-1), 5.5 (m, H-4), 5.3 and 5.1 (m, H-3 and -2), 4.4 (t, H-5), 3.5 and 3.4 (m, H-6 and - $\alpha$ ), 2.0, 2.05 and 2.1 (s, COCH<sub>3</sub>), 1.2 [m, (CH<sub>2</sub>)<sub>6</sub>], 0.9 (t, CH<sub>3</sub>). The bromide **8** was immediately used for glycosylation reactions.

1,2:3,4-Di-O-isopropylidene-6-O-(2,3,4-tri-O-acetyl-6-O-octyl-α-D-galactopyranosyl)- $\alpha$ -D-galactopyranose (9). — To a solution of the bromide 8 (1.28 g, 2.66 mmol) in dichloromethane (15 mL) at  $-15^{\circ}$  under an atmosphere of Ar, were added silver triflate (0.7 g, 2.7 mmol) and a solution of 1,2:3,4-di-O-isopropylidene- $\alpha$ -D-galactopyranose (0.7 g, 2.66 mmol) and N,N,N',N'-tetramethylurea (0.31 g, 2.66 mmol) in dichloromethane (10 mL). After 20 min at  $-15^{\circ}$ , the mixture was poured into a cold solution of NaHCO3 and extracted with dichloromethane, washed with water, and dried (MgSO<sub>4</sub>). The crude product was applied to a silica gel column and eluted with 1:1 diethyl ether-pentane to give  $9 (0.72 \text{ g}, 40\%), [\alpha]_D^{20}$  $-46^{\circ}$ ,  $[\alpha]_{546}^{20}$  -54° (c 1.6, methanol); <sup>1</sup>H-n.m.r.:  $\delta$  5.50 (d,  $J_{1.2}$  4.9 Hz, H-1), 5.43  $(q, J_{4',5'} 1.1 \text{ Hz}, H-4'), 5.21 (q, J_{2',3'} 10.5 \text{ Hz}, H-2'), 5.02 (q, J_{3',4'} 3.4 \text{ Hz}, H-3'),$ 4.58 (q,  $J_{3,4}$  7.9 Hz, H-3), 4.56 (d,  $J_{1',2'}$  8.0 Hz, H-1'), 4.29 (q,  $J_{2,3}$  2.4 Hz, H-2), 4.18 (q,  $J_{4.5}$  1.9 Hz, H-4), 4.05 (q,  $J_{6A.6B}$  11.5 Hz, H-6A), 3.93 (m,  $J_{5.6A}$  3.3,  $J_{5.6B}$ 7.6 Hz, H-5), 3.81 (m,  $J_{5',6'A}$  6.0,  $J_{5',6'B}$  6.9 Hz, H-5'), 3.67 (q, H-6B), 3.54 (q,  $J_{6'A.6'B}$  9.8 Hz, H-6'A), 3.44 (q, H-6'B), 3.43 (m, H- $\alpha$ A), 3.34 (m, H- $\alpha$ B), 2.13, 2.08, 1.98 (3 s, OAc), 1.51, 1.44, 1.32 [3 s, C(CH<sub>3</sub>)<sub>2</sub>], 1.26 [m, (CH<sub>2</sub>)<sub>6</sub>], and 0.88 (t, CCH<sub>3</sub>)<sub>2</sub>]CH<sub>3</sub>);  ${}^{13}$ C-n.m.r.:  $\delta$  171.8, 171.5 (C=O), 110.5, 109.4 (O-C-O), 103.1 (C-1'), 97.7 (C-1), 72.9–62.9 (C cyclic), and 32.9–14.4 (remaining C).

Anal. Calc. for C<sub>32</sub>H<sub>52</sub>O<sub>14</sub>: C, 58.1; H, 7.9. Found: C, 57.9; H, 7.8.

6-O-(6-O-Octyl- $\alpha$ -D-galactopyranosyl)-D-galactopyranose (10). — Compound 9 (690 mg) was dissolved in 90% aqueous trifluoroacetic acid (5 mL). The solution was kept 1 h at 0°, and then evaporated to give a chromatographically pure compound which was deacetylated in methanol (10 mL) with triethylamine (3 mL) for 24 h at room temperature. The solution was evaporated and the residue crystallized from methanol—ethyl acetate to give 10, m.p. 129°,  $[\alpha]_D^{20} + 7.5^\circ$ ,  $[\alpha]_{546}^{20} + 8.6^\circ$  (c 1, methanol); <sup>1</sup>H-n.m.r.  $[(^2H_4)$ -methanol]:  $\delta$  5.17 (H-1), 4.5–3.2 (H, cyclic), 1.32 [m, (CH<sub>2</sub>)<sub>6</sub>], and 0.9 (t, CH<sub>3</sub>).

Anal. Calc. for C<sub>20</sub>H<sub>38</sub>O<sub>11</sub>: C, 52.8; H, 8.4. Found: C, 52.7; H, 8.3.

## **ACKNOWLEDGMENTS**

The authors thank C. Vincent for helpful discussions and the Ministère de l'Industrie et de la Recherche (Essor des Biotechnologies) for financial support.

#### REFERENCES

- 1 A. ZAKS AND A. M. KLIBANOV, Proc. Natl. Acad. Sci. U.S.A., 82 (1985) 3192-3196.
- 2 A. MATSUSHIMA, M. OKADA, AND Y. INADA, FEBS Lett., 178 (1984) 275-277.
- 3 T. J. AKERN AND A. M. KLIBANOV, Science, 228 (1985) 1280-1284.
- 4 G. D. KUTUZOVA, N. N. UGAROVA. AND I. V. BEREZIN, Russ. Chem. Rev., (Engl. Transl.), 53 (1984) 1078–1100.
- 5 R. D. SCHMID, Adv. Biochem. Eng., 12 (1979) 41-118.
- 6 K. MARTINEK AND I. V. BEREZIN, J. Solid Phase Biochem., 2 (1977) 343-385.
- 7 T. ARAKAWA AND S. N. TIMASHEFF, Biochemistry, 21 (1982) 6536-6544.
- 8 G. W. STUBBS, H. G. SMITH, AND B. J. LITMAN, Biochim. Biophys. Acta 426 (1976) 46-56.
- 9 J. E. K. HILDRETH, Biochem. J., 207 (1982) 363-366.
- 10 M. H. REMY AND D. THOMAS, Enzyme Microbiol. Technol., 4 (1982) 381-384.
- 11 C. P. STOWELL AND Y. C. LEE, Adv. Carbohydr. Chem. Biochem., 37 (1980) 225-281.
- 12 H. PAULSEN, Angew. Chem., Int. Ed. Engl., 21 (1982) 155-224.
- 13 K. IGARASHI, Adv. Carbohydr. Chem. Biochem., 34 (1977) 243-283.
- 14 M. G. VAFINA AND N. V. MOLODTSOV, Carbohydr. Res., 47 (1976) 188-194.
- 15 A. YA, KHORLIN, M. L. SHUL'MAN, S. E. ZURABYAN, I. M. PRIVALOVA. AND Y. L. KOPAEVICH, Izv. Akad. Nauk SSSR, Ser. Khim., (1968) 2094–2098.
- 16 R. U. LEMIEUX AND H. DRIGUEZ, J. Am. Chem. Soc., 97 (1975) 4063-4068.
- 17 J. E. CHRISTENSEN AND L. GOODMAN, Carbohydr. Res., 7 (1968) 510-512.
- 18 J. S. Brimacombe, Methods Carbohydr. Chem., 6 (1972) 376-378.
- 19 B. HAVLINOVA, M. KOSIK, P. KOVÁČ, AND A. BLAZEJ, Tenside Detergents, 15 (1978) 72-74.
- 20 S. HANESSIAN AND J. BANOUB, Carbohydr. Res., 53 (1977) c13-c16.
- 21 S. L. PATT. J. Carbohydr. Chem., 3 (1984) 493-511.